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Effect of Prebiotic (Fermacto) in Low Protein Diet on Performance and Carcass Characteristics of Broiler Chicks

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Abstract: This study was conducted to evaluate the effects of prebiotic (Fermacto) in low protein diet on performance and carcass characteristics of broiler chicks. One hundred and fifty six 1-day old Ross 308 broiler chicks of both sexes were used for 42-days. The chicks were randomly allocated to 12 pens containing 13 chicks each with 3 replicates and assigned to receive one of the 4 dietary treatments of 2 levels of protein (low and high) and 2 levels of prebiotic (0 and 0.2%) in a completely randomized design with factorial arrangement. There was not significant difference in feed conversation ratio, Feed intake and tights weight among treatments. Significant differences were observed in carcass weight and abdominal fat percentage between high protein diet without prebiotic and low protein diet containing prebiotic (p<0.05). The results of the present experiment showed that addition of prebiotic to broiler diets containing 90% of NRC protein recommendation had same effect with control diet on performance of broiler chicks.

Key words: Prebiotic, low protein diet, performance, broiler chicks

Introduction

Starting in 2006 the prophylactic use of antibiotics or so called Antimicrobial Growth Promoters (AGP'S) in animal feeds will be banned in the EU. These AGP'S are use to improve the feed efficiency by altering the microflora in the Gastro Intestinal Tract (GIT). The main reason for this ban is the increasing resistance of pathogenic bacteria against antibiotics.

Use of prebiotics or fermentable sugars instead of antibiotics is going to be popular in birds in order to improve the useful microbial population of GIT (Kermanshahi and Rostami, 2006). Prebiotics have been defined by Gibson and Roberfroid (1995) as indigestible food ingredients which stimulate the growth and/or activity of a select number of bacteria in the GIT and improve the host's health. Prebiotics have been shown to alter gastrointestinal microflora, alter the immune system, prevent colonic cancer, reduce pathogen invasion including pathogens such as Salmonella Entritidis and E.coli and reduce cholesterol and odor compounds (Cummings and Macfarlane, 2002; Cummings et al., 2001; Simmering and Blaut, 2001). The population of useful bacteria likes lactobacillus and bifidobacteria (Ziggers, increases and the pH of the GIT, due to increasing production of volatile fatty acids, decreases. Therefore the environment of GIT becomes unsuitable for the activity and proliferation of pathogens like Salmonella. Based on Nurmi concept of competitive exclusion (Nurmi and Rantala, 1973), pathogens will be expelled out of the gut by useful bacteria if it already occupied the gut sites.

Colonization of useful micro flora in the gut of young birds that is related to gut conditions can inhibit further colonization of pathogens (Nurmi and Rantala, 1973). High protein prices and environmental concerns have pressured the industry to reduce dietary protein levels (Firman, 1997). Thus, low protein diets are of interest and important for feed additive evaluation and animal performance. The commercially available fermentation product of Aspergillus orizae, Fermacto (PET-AG, 2006), referred to as Aspergillus meal (AM), has no live cells or spores (PET-AG, 2006) and is proven to enhance the digestive efficiency of the gut (Harms and Miles, 1988). Aspergillus meal might offer better results when the level of protein and amino acids is lower than those recommended by NRC (1994) or applied in commercial flocks. Lower levels of protein and amino acids as compared with those recommended by NRC (1994) or applied in commercial flocks are also a potential environmental benefit (Torres-Rodriguez et al., 2005). Because of reports on the use of AM and low dietary protein and amino acids on performance are lacking, Therefore, the objective of the present study was to evaluate the effect of prebiotic (Fermacto) in low protein diet on performance and carcass characteristics of broiler chicks.

Materials and Methods

Birds and experimental diets: 156 day-old mixed Ross broiler chicks were randomly allocated to 12 groups of 13 birds each and reared for 42 days. There were four treatments (treatment 1: high protein diet without prebiotic, treatment 2: high protein diet containing

Table 1: Composition of experimental diets in different periods of the experiment

	Starter		Grower		Finisher	
Ingredient (%)	Low Protein	HighProtein	Low Protein	HighProtein	Low Protein	HighProtein
Corn	66.50	59.10	71.30	64.46	76.09	69.92
Soybean meal	29.10	35.50	24.02	29.64	19.63	24.72
DCP	1.74	1.66	1.62	1.57	1.53	1.48
Calcium carbo.	1.20	1.14	1.08	1.07	1.03	1.02
Premix	0.50	0.50	0.50	0.50	0.50	0.50
Salt	0.30	0.30	0.30	0.30	0.30	0.30
Sunflower oil	-	1.10	0.72	1.97	0.65	1.79
DL-Methionine	0.28	0.34	0.18	0.23	0.10	0.14
L-Lysine	0.38	0.36	0.28	0.26	0.17	0.13
Composition						
Metab. Energy (Kcal/Kg)	2900	2900	3000	3000	3050	3050
Crude Protein (%)	19.00	21.20	17.00	18.90	15.32	17.02
Calcium (%)	0.90	0.90	0.85	0.85	0.80	0.80
Avail. Phos. (%)	0.45	0.45	0.42	0.42	0.40	0.40
Arginine (%)	1.17	1.34	1.02	1.17	0.91	1.04
Lysine (%)	1.24	1.38	1.04	1.16	0.85	0.94
Met+Cys (%)	0.94	1.05	0.80	0.90	0.68	0.76
Sodium (%)	0.13	0.13	0.13	0.13	0.13	0.13

Table 2: Effect of protein levels on feed intake of broiler chicks

	Feed i	ntake (g)				
Treatment	1-10	10-28		28-	42	1-42
High protein	267°	1595°		267	' 5	4537
Low protein	253₺	1432b		251	1	4197
SEM	0.003	0.030		0.08	38	0.107
Meane with	different	cupercorinte	in	each	column	diffor

Means with different superscripts in each column different significantly (p<0.05)

Table 3: Effect of prebiotic levels on feed intake of broiler chicks

	Feed int	Feed intake (g)		
Treatment	1-10	10-28	28-42	1-42
Diet without prebiotic	260	1540	2621	4420
Diet contain 0.2% prebiotic	261	1488	2565	4314
SEM	0.003	0.030	0.088	0.107
Means with different so	uperscripts	in each	column	differ
significantly (p<0.05)				

Table 4: Effect of protein and prebiotic levels on feed intake of broiler chicks

	Feed intake (g)			
Treatment	1-10	10-28	28-42	1-42
High protein without prebiotic	266ª	1602ª	2650	4520
High protein with prebiotic	267ª	1588°	2700	4555
Low protein without prebiotic	252b	1476ab	2593	4321
Low protein with prebiotic	254ªb	1389⁵	2429	4073
SEM	0.004	0.043	0.124	0.151
Means with different super significantly (p<0.05)	scripts	in each	column	differ

prebiotic, treatment 3: low protein diet without prebiotic, treatment 4: low protein diet containing prebiotic) in this experiment. Prebiotic (Fermacto) was supplemented at the rate of 0 and 2.0 kg/ton of diets. Feed and water were provided *ad libitum* during the experiment. Diets were provided in 3 periods of starter (1-10 days), grower

(10-28 days) and finisher (28-42 days) of age. Composition of experimental diets is presented in Table 1

Data and sample collection: Body weight gain, feed intake and feed conversion ratio were recorded weekly. Mortality was recorded throughout of the experiment. At 42 days of age two birds from each replicate with body weight similar to mean replicate body weight were selected, slaughtered and the fat pad, carcass, breast meat and tights weight were measured.

Data analysis: The data from this experiment were subjected to one-way analysis of variance as factorial arrangement 2×2 with 2 levels of protein and 2 levels of prebiotic, thus there was 4 treatments and 3 replicates for each treatment. The obtained data were submitted to analysis of variance, using the General Linear Model procedure (GLM) of SAS software (SAS Institute, 2002). Means were compared by the Duncan's multiple range tests at 5% probability (Duncan, 1955).

Results and Discussion

Effects of different levels of protein and prebiotics on feed intake of broiler chicks in different phases of the experiment are presented in Table 2 and 3. There was significant differences among treatments for feed intake in starter and grower periods of the experiment for protein levels (p<0.05). chicks fed with high protein diets consumed more feed in starter and grower phase of the experiment. Effect of prebiotic on feed intake of chicks in all phases of the experiment was not significant. Interaction of protein and prebiotic are shown in Table 4. Interaction of protein and prebiotic on feed intake was significant in starter and grower periods of the experiment (p<0.05).

Table 5: Effect of protein levels on body weight gain of broiler

	CHICK	15					
		Body v	Body weight gain (g)				
Treatme	nt	1-10	10-28		28-	42	1-42
High pro	tein	198ª	983°		132	21	2502
Low prof	tein	183 ^b	888 ^b		124	2	2313
SEM		0.003	0.016		0.0	30	0.042
Means significa	with ntly (p<	different (0.05)	superscripts	in	each	column	differ

Table 6: Effect of prebiotic levels on body weight gain of broiler

CHICKS						
	Body weight gain (g)					
Treatment	1-10	10-28	28-42	1-42		
Diet without prebiotic	189	934	1286	2410		
Diet contain 0.2% prebiotic	191	936	1277	2405		
SEM	0.003	0.016	0.030	0.042		

Means with different superscripts in each column differ significantly (p<0.05)

Table 7: Effect of protein and prebiotic levels on body weight gain of broiler chicks

gain of broller critical	,			
	Body weight gain (g)			
Treatment	1-10	10-28	28-42	1-42
High protein without prebiotic	197ªb	1005°	1323	2525°
High protein with prebiotic	199ª	961ab	1319	2479ab
Low protein without prebiotic	181⁰	865⁰	1250	2296b
Low protein with prebiotic	185 ^{bc}	911bc	1235	2331ab
SEM	0.004	0.023	0.043	0.060
Manua with different arms		in anala		4:45

Means with different superscripts in each column differ significantly (p<0.05)

Table 8: Effect of protein levels on feed conversion ratio of broiler chicks

	Feed conversion ratio						
Treatment	1-10	10-28	28-42	1-42			
High protein	1.350	1.625	2.027	1.815			
Low protein	1.386	1.615	2.018	1.814			
SEM	0.017	0.034	0.054	0.040			

Means with different superscripts in each column differ significantly (p<0.05)

Table 9: Effect of prebiotic levels on feed conversion ratio of broiler chicks

	Feed conversion ratio				
Treatment	1-10	10-28	28-42	1-42	
Diet without prebiotic	1.375	1.652	2.039	1.836	
Diet contain 0.2% prebiotic	1.361	1.590	2.006	1.793	
SEM	0.017	0.034	0.054	0.040	

Means with different superscripts in each column differ significantly (p<0.05)

Effect of protein and prebiotic levels on body weight gain are presented in Table 5 and 6. Effect of protein levels on body weight gain was significant in starter and grower periods (p<0.05). Interaction of above 2 factors was significant in starter, grower and whole phases of

Table 10: Effect of protein and prebiotic levels on feed conversion ratio of broiler chicks

	Feed co	Feed conversion ratio				
Treatment	1-10	10-28	28-42	1-42		
High protein without prebiotic	1.356	1.595⁴⁵	2.007	1.792		
High protein with prebiotic	1.344	1.654ª⁵	2.046	1.838		
Low protein without prebiotic	1.394	1.708°	2.070	1.880		
Low protein with prebiotic	1.379	1.523⁵	1.967	1.747		
SEM	0.025	0.048	0.076	0.056		

Means with different superscripts in each column differ significantly (p<0.05)

Table 11: Effect of protein levels on carcass characteristics of broiler chicks

	Carcass characteristics					
Treatment	Carcass (%)	Breast (%)	Tights (%)	 Abdominal fat (%)		
High protein	79.235	27.857ª	22.544	3521		
Low protein	77.875	25.680⁵	23.124	3.430		
SEM	0.434	0.431	0.338	0.184		

Means with different superscripts in each column differ significantly (p<0.05)

the experiment (P < 0.05). Results are shown in Table 7. Commercial diets (with and without addition of AM) caused heavier body weights than the 2 low protein diets, however, addition of AM to low protein diet improved body weight in starter, grower and total periods of the experiment. Differences between the commercial and low protein diets were expected because lower protein and amino acids (lysine, methionine and threonine) content affect weight gain and performance. The difference observed between the low protein and low protein supplemented with prebiotic can be attributed to the effect of prebiotic, because these 2 diets were formulated to supply the same amount of protein and amino acids. Effect of protein and prebiotic level on feed conversion ratio are presented in Table 8, 9, 10. Interaction of 2 factors In grower periods, was significant for this trait (p<0.05). Treatment 4 (low protein with periodic) had the lowest and treatment 3 had the highest value for this trait.

Effects of experimental diets on carcass characteristics are presented in Table 11, 12 and 13. Chicks fed with high protein diets had more breast meat percentage (p<0.05). There was significant difference among groups for carcass and breast meat percentage (p<0.05). Carcass and breast meat percentage in low protein with prebiotic was lower than other groups. Breast meat percentage in treatment 1 was higher than treatment 3. There were not significant differences among treatments for tights and abdominal fat percentage.

Prebiotics have been shown to alter gastrointestinal microflora, alter the immune system, prevent colonic cancer, reduce pathogen invasion including pathogens such as *E.coli* and reduce cholesterol and odor compounds (Cummings and Macfarlane, 2002; Cummings *et al.*, 2001; Simmering and Blaut, 2001).

Table 12: Effect of prebiotic levels on carcass characteristics of broiler chicks

·	Carcass characteristi	Carcass characteristics					
Treatment	Carcass (%)	Breast (%)	 Tights (%)	Abdominal fat (%)			
Diet without prebiotic	79.143	26.354	22.606	3.531			
Diet contain 0.2% prebiotic	77.966	27.182	23.061	3.425			
SEM	0.434	0.431	0.338	0.184			

Means with different superscripts in each column differ significantly (p<0.05)

Table 13: Effect of protein and prebiotic levels on carcass characteristics of broiler chicks

Treatment	Carcass characteristics			
	Carcass (%)	Breast (%)	 Tights (%)	Abdominal fat (%)
High protein without prebiotic	79.338°	27.451 ^{ab}	22.107	3.570
High protein with prebiotic	79.142°	28.262°	22.980	3.330
Low protein without prebiotic	78.960°	25.258°	23.105	3.337
Low protein with prebiotic	76.790 ^b	26.102 ^{bc}	23.143	3.521
SEM	0.613	0.610	0.478	0.260

Means with different superscripts in each column differ significantly (p<0.05)

Prebiotics are short-chain carbohydrates that are indigestible by human, animal and poultry digestive systems. Prebiotics stimulate the growth and/or activity of a select number of bacteria in the GIT and improve the host's health. (Cummings and Macfarlane, 2002; Cummings et al., 2001; Patterson and Burkholder, 2003). The major effects of prebiotics have been reviewed by Cummings and Macfarlane (2002) and include: production of short-chain fatty acids and lactate, selective increases in bifidobacteria and lactobacilli, increase in pathogen resistance and improved calcium and magnesium absorption. The results observed for body weight gain are in accordance with other reports (Meinz, 1993; Tangendjaja, 1993) that supplementation of Fermacto in broiler diet resulted in weight gain than those using diet without supplemented of Fermacto. These findings are also consistent with reports of improved body weight gain when broilers are fed diet supplemented with prebiotics (Ammerman and Twining, 1989; Yusrizal and Chen, 2003). Several studies have shown that addition of prebiotics to the diet of broiler, layer and pig leads to improved performance through improving gut microflora (Pelicano et al., 2004; Spring et al., 2000; Xu et al., 2003). Zhang (2000) reported that 0.2 or 0.4% Isomaltooligosaccharide (IMO) (a prebiotic) could improve broiler performance in the starter period. Recent report suggested that feeding of chicory beta fructans-an oligosaccharide, reduced the abdominal fat of broiler chicken (Yusrizal and Chen, 2003). In recent study, Kalavathy et al. (2003) found that supplementation of Lactobacillus culture significantly reduced abdominal fat. Similarly, Yusrizal and chen (2003) reported that supplementation of beta fructans from chicory had significantly produced low level of abdominal fat. Our findings are contrast with the results obtained from mentioned studies. Others have noted a lack of change in the size of the fat pad after a similar period of feeding (Waldroup et al., 1993). Xia et al. (2001) reported that

dietary IMO improved survivability of chicks. These results are same with our results. It is probable that IMO supplementation altered the gastrointestinal microflora and, therefore, affected the immune system (Li *et al.*, 2000; Shao *et al.*, 2000). The results of the present study indicated that addition of prebiotic to broiler diets containing 90% of NRC protein recommendation (low protein diet) had similar effect on performance like control diet (high protein diet without prebiotic).

References

Ammerman, E., C. Quarles and P.V. Twining, 1989. Evaluation of fructooligosaccharides on performance and carcass yield of male broiler. Poult. Sci., 68 (Suppl.), 167.

Cummings, J.H. and G.T. Macfarlane, 2002. Gastrointestional effects of prebiotics. Br. J. Nutr., 87 (Suppl. 2): 145-151.

Cummings, J.H., G.T. Macfarlane and H.N. Englyst, 2001. Prebiotic digestion and fermentation. Am. J. Clin. Nutr., 73 (Suppl): 415S-420S.

Duncan, D.B., 1955. Multiple ranges and multiple F. test. Biometric, 11: 1-42.

Firman, J.D., 1997. Early stage turkey nutrition and ideal protein: Implications for all aspects of poultry production. Pages 57-67 in Nutritional Biotechnology in the Feed and Food Industries. Proc. Alltech's 20th Symp. T.P. Lyons and K.A. Jacques, Ed. Nottinham University Press, UK.

Gibson, G.R. and B. Roberfroid, 1995. Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. J. Nutr., 125: 1401-1412.

Harms, R.H. and R.D. Miles, 1988. Research note: Influence of Fermacto on the performance of laying hens when fed with different levels of methionine. Poult. Sci., 67: 842-844.

- Kalavathy, R., N. Abdullah, S. Jalaludin and Y.W. Ho, 2003. Effect of Lactobacillus cultures on growth performance, abdominal fat deposition, serum lipids and weight of organs of brother chickens. Br. Poult. Sci., 44: 139-144.
- Kermanshahi, H. and H. Rostami, 2006. Influence of supplemental dried whey on broiler performance and cecal flora. Int. J. Poult. Sci., 5: 538-543.
- Li, M., G.Y. Zhao, G.C. Zhao and Q.X. Gao, 2000. Effect of isomalto-oligosaccharides on cellular immunity of piglets. Feed Res., 11: 7-9.
- Meinz, J., 1993. 100 ppm Monensin verses 0.2% Fermacto. Fermacto, pet. Ag. Inc Elgin, Illinois 60120, USA.
- National Research Council, 1994. Nutrient Requirements of Poultry. 9th edition (Revised). National Academy Press. Washington, D.C.
- Nurmi, E. and M. Rantala, 1973. New aspects of Salmonella infection in broiler production. Nature, 241: 210-211.
- Patterson, J.A. and K.M. Burkholder, 2003. Application of prebiotics and prebiotics in poultry production. Poult. Sci., 82: 627-631.
- Pelicano, E.R.L., P.A. De Souza, H.B.A. De Souza, F.R. Leonel, N.M.B.L. Zeola and M.M. Boiago, 2004. Productive traits of broiler chickens fed diets containing different growth promoters. In: Asian Tec. Conf. Brazil, pp: 18.
- PET-AG, 2006. A subsidiary of Borden, Inc., Elgin, IL.
- SAS Institute, 2002. SAS Uers Guide Statics. Version 8.1 Ed. SAS institute Inc., Cary, NC.
- Shao, L.P., L.J. Zhou, Z.F. Fei, G.P. Li, F.P. Lin and T. Jin, 2000. Effect of mannan-oligosaccharide on immune function and blood antioxidant enzymes in sucking piglets. Acta Nutr. Sin., 1: 82-84.
- Simmering, R. and M. Blaut, 2001. Pro- and Prebioticsthe tasty guardian angles? Appl. Microbiol. Biotechnol., 55: 19-28.

- Spring, P., C. Wenk, K.A. Dawson and K.E. Newman, 2000. The effects of dietary mannan oligosaccharides on cecal parameters and the concentrations of enteric bacteria in the caeca of *Salmonella*-challenged broiler chicks. Poult. Sci., 79: 205-211.
- Tangendjaja, B., 1993. Effect of Fermacto upon the utilization of broiler diets containing normal and high levels of rice bran. Ilmu Peternakan, 7: 16-19.
- Torres-Rodriguez, A., C. Sartor, S.E. Higgins, A.D. Wolfenden, L.R. Bielke, C.M. Pixley, L. Sutton, G. Tellez and B.M. Hargis, 2005. Effect of Aspergillus meal prebiotic (Fermacto) on performance of broiler chickens in the starter phase and fed low protein diets. J. Appl. Poult. Res., 14: 665-669.
- Waldroup, A.L., J.T. Skinner, R.E. Hierholzer and P.W. Waldroup, 1993. An evaluation of fructooligosaccharides in diets for broiler chickens and effects on Salmonellae contamination of carcasses. Poult. Sci., 72: 643-650.
- Xia, Z.S., J.H. Xia, Y.H. Quan and J.L. Zhou, 2001. The effect of dietary with isomalto-oligosaccharides on the performance of yellow broiler chicks. Cereal Feed Ind., 8: 32-33.
- Xu, Z.R., C.H. Hu, M.S. Xia, X.A. Zhan and M.Q. Wang, 2003. Effects of dietary fructo oligosaccharide on digestive enzyme activities. Intestinal Microflora and morphology of male broilers. J. Anim. Sci., 82: 1030-1036.
- Yusrizal and T.C. Chen, 2003. Effect of adding chicory fructons in feed on broiler growth performance, serum cholesterol and intestinal length. Int. J. Poult. Sci., 2: 214-219.
- Zhang, C., 2000. Research on the application effects and the mechanism of isomalto-oligosaccharides in diet for broilers and piglets. Treatise. China Agricultural University, Beijing.
- Ziggers, D., 2000. Tos, a new prebiotic drived from whey. Anim. Feed Sci. and Tech., 5: 34-36.