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Research Article

The Effect of Different Extenders on Some Fertility Properties of Roosters Semen

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Abstract

Background and Objective: Poultry sperm gradually loses its quality within an hour of sample collection. This study aimed to determine the storage effects of different extenders (at 4 °C) on rooster sperm fertility properties. **Materials and Methods:** Eighteen seminal fluid samples of a Lohman Brown strain were collected by dorsal-abdominal massage. These were divided into three groups and added to three extenders: a simple medium for assisted reproductive technology (SMART), Tris and milk. Since non-diluted fresh sperm loses its quality within an hour of semen collection, no control group was used. Percentages of mass activity, motility, normality and viability were tested. These were done at 1, 4 and 8 h after semen collection. Completely Random Design (C.R.D) was used in this experiment. **Results:** At the first timepoint, the semen test score of the SMART medium extender was higher than that of the other extenders. In contrast, the milk extender shows a highly significant improvement in sperm parameters post preservation at both the second and the third time tests. The SMART extender was protecting sperm vigor more than Tris at most of the tested timepoints. **Conclusion:** Extenders delay the loss of rooster sperm fertilization ability. Full milk extender was better than other extenders in the protection of sperm fertility.

Key words: Mass activity, milk extender, rooster sperm, semen motility, semen viability

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Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Poultry semen is highly concentrated (billions of sperms per ml) viscous fluid. Therefore, it needs to be diluted before being used for artificial insemination¹. Semen dilutions improve the processing of semen tests such as their concentration, motility, abnormality, vitality and pH². Extender solutions maintain the functional proprieties of sperms and increasing the semen volume³. The optimal osmotic pressure for rooster semen extender is between 325 and 350 mOsmol kg⁻¹⁴. Artificial insemination success depends on the efficiency of semen collection and storage⁵. Artificial insemination is better than the natural mating for broiler breeder parents stock⁶. Sperms storage of chickens and turkeys without dilution is inefficient processes⁷. As the dilution ratio increase, the negative effect on sperm motility in guinea fowls increases, as well⁸. A dilution rate of 1/10 was shown to affects sperm quality, negatively⁹. Extenders with different ingredient are used to sustain sperm vitality, such as glycerol and egg yolk¹⁰, lactated Ringer's glucose, glucose-Tris-glucose and lactated Ringer's extenders¹¹, skim milk and Tris-citrate extenders¹², orange juice¹³ and vitamins of A, C and E¹⁴. This study was conducted to determine the efficiency of the understudy extenders in preserving rooster semen under short-term storage (1, 4 and 8 h). These were done by evaluating physical semen proprieties.

MATERIALS AND METHODS

This experiment was conducted in the College of Agriculture-the University of Al-Qadisiyah, Iraq. Eighteen Lohman Brown roosters 1-year-old were used. The roosters were divided randomly into three groups and trained for having semen. It collected by dorsal-abdominal message two weeks in advance. Extenders were assigned randomly to each semen group. Three extenders were used:

SMART extender¹⁵: A ringer solution composed of 29 mmol L⁻¹ bicarbonate, 3.2 g sodium lactate, 6.0 g sodium chloride, 0.4 g potassium chloride and 0.27 g calcium chloride

per liter of distilled water. Specific chemicals were applied to this solution, including 0.5 g of red phenol, 0.01 g of sodium pyruvate and human serum albumin (HSA, 20%) and stored in a special non-toxic bottle.

Tris extender¹⁶: The semen diluent was prepared by dissolving 3.8 g Tris, 2.2 g citric acid and 0.6 g glucose into 100 mL of distilled water.

Full cream milk¹⁷: Every 100 mL of full cream milk contains 3.2 g total fat (saturated fat of 2.17 g and unsaturated fat of 1.03 g), 7.53 mg cholesterol, 5.4 g lactose, 3.2 g protein, 0.01 g dietary fiber and 0.05 g sodium.

The dilution ratio was 1:3 (1 semen: 3 extenders). Dilutions were sterilized with streptomycin (100 mg) and penicillin (100,000 IU). Data were taken at 1, 4 and 8 h after semen collection.

Evaluation of semen: Mass activity, sperm motility, sperm viability and normality percentage were estimated according to Blom and Christensen¹⁸, Chemineau *et al.*¹⁹, Hancock²⁰ and Swanson and Bearden²¹ methods, respectively.

Statistical analysis was performed using the SPSS18 software according to Completely Random Design (C.R.D). The Duncan multiple ranges test was used to compare differences among means. Significance levels of $p \leq 0.05$ and $p \leq 0.01$ were used.

RESULTS

The result showed the effect of different extenders after an hour of semen collection (Table 1). The SMART extender scored significantly higher in mass activity and sperm viability tests compared to the milk extender ($p \leq 0.05$). Also, the SMART medium outperformed the Tris and milk extenders in the individual motility test. The Tris extender had significantly higher values more than the milk dilution for the same test. This shows non-significant differences among the extenders at $p \leq 0.05$, in accordance with the sperm normality (%) test. Highly significant differences among extenders at ($p \leq 0.01$)

Table 1: The effect of tested extenders on rooster sperm parameters at 1 h after collection

Parameters	SMART medium	Tris	Milk	p-value
Mass activity (%)	92.667 ± 1.45 ^A	90.667 ± 0.67 ^{AB}	88.333 ± 1.67 ^B	0.1500NS
Individual motility (%)	88.333 ± 1.67 ^A	82.333 ± 1.45 ^B	79.000 ± 1.00 ^C	0.0090**
Sperm normality (%)	93.333 ± 1.67 ^A	92.333 ± 1.45 ^A	91.667 ± 1.67 ^A	0.7684NS
Sperm viability (%)	94.333 ± 0.67 ^A	90.000 ± 2.89 ^{AB}	87.000 ± 1.53 ^B	0.0910NS

Different letters denote to significant differences at $p \leq 0.05$. Similar letters denote to non-significant differences at $p > 0.05$, **High significant different ($p \leq 0.01$), NS: No significant differences ($p > 0.05$)

Table 2: The effect of tested extenders on rooster sperm parameters at 4 h after collection

Parameters	SMART medium	Tris	Milk	p-value
Mass activity (%)	70.667±0.67 ^C	63.667±0.88 ^B	82.667±1.45 ^A	0.0006**
Individual motility (%)	66.333±0.88 ^B	59.333±0.67 ^C	74.333±2.33 ^A	0.0012**
Sperm normality (%)	83.333±1.67 ^B	74.667±1.45 ^C	88.000±1.53 ^A	0.0025**
Sperm viability (%)	76.667±0.88 ^B	63.000±1.53 ^C	81.000±1.00 ^A	0.0001**

Different letters denote to significant differences at $p \leq 0.05$. Similar letters denote to non-significant differences at $p > 0.05$, **High significant different ($p \leq 0.01$)

Table 3: The effect of tested extenders on rooster sperm parameters at 8 h after collection

Parameters	SMART medium	Tris	Milk	p-value
Mass activity (%)	62.000±2.00 ^A	41.667±0.88 ^B	66.667±1.67 ^A	0.0001**
Individual motility (%)	57.667±1.45 ^B	37.667±1.45 ^C	62.667±2.67 ^A	0.0002**
Sperm normality (%)	53.333±1.67 ^B	42.000±1.15 ^C	61.667±1.67 ^A	0.0003**
Sperm viability (%)	60.667±0.67 ^B	45.667±1.20 ^C	74.667±1.33 ^A	0.0008**

Different letters denote significant differences at $p \leq 0.05$, Similar letters denote non-significant differences at $p > 0.05$. **High significant different ($p \leq 0.01$)

only in the individual motility test was observed. After 4 h of semen collection, the milk extender outperforms the other extenders at protecting sperm (Table 2, $p \leq 0.01$). At this test time, the SMART outperformed the Tris extender in the individual motility, sperm viability and sperm normality tests. In contrast, the Tris extender outperformed the SMART medium extender in the mass activity test. Full milk dilator had significantly higher values ($p \leq 0.01$) as compared to the other extenders. The SMART medium extender significantly outperforms the Tris extender in all tests (Table 3).

DISCUSSION

Extenders delay may cause loss of rooster sperm fertilization ability but the tests values decrease as time increases. This is consistent with the findings of Hudson *et al.*⁸, who reported that the storage period at 5°C has a significant influence on sperm motility in guinea fowls. Sperm motility and viability gradually decline after collection¹². SMART medium performed best at 1 h after collection. This is because the SMART medium includes serum albumin, which acts as an antioxidant and pyruvate, which acts as an energy source^{22,23}. However, at 4 or 8 h after collection, the full milk extender outperformed the other extenders. This is in line with the findings of Rahman, who reported that milk dilution significantly increases sperms viability as compared to the Tris extender in both viability (%) and sperm motility (%) tested at different periods in ram¹². This might be because the sperm contains low cholesterol, phospholipid and low protein phospholipid²⁴. Fatty acids affect membrane liquidity^{25,26}. The cholesterol and fatty acid contents increase in the plasma membranes that have a low percentage, causes the membranes to become more resistant to oxidative damage that resulted in increasing protection. Milk casein decreases damage to cell membrane lipids and improves sperms motility and viability^{27,28}.

CONCLUSION

The poultry industry is always searching for new sublimates and extenders to improve the efficiency of prolonging sperms life and activity. This study examined the storage effects of different extenders on roosters sperm fertility properties. Extenders delay the deficiency of roosters sperms fertilization ability. Birds semen dilutions make work with poultry breeding much easier and allow for the insemination a large number of females. The inclusion of milk dilator better protects sperm fertility compared to the SMART and Tris extenders.

REFERENCES

1. Donoghue, A.M. and G.J. Wishart, 2000. Storage of poultry semen. *Anim. Reprod. Sci.*, 62: 213-232.
2. Alkan, S., A. Baran, O.B. Ozdas and M. Evecen, 2002. Morphological defects in turkey semen. *Turk. J. Vet. Anim. Sci.*, 26: 1087-1092.
3. Foote, R.H., 2002. Within-herd use of boar semen at 5°C, with a note on electronic monitoring of oestrus. *Reprod. Domest. Anim.*, 37: 61-63.
4. Sexton, T.J. and T.A. Fewlass, 1978. A new poultry semen extender: 2. Effect of the diluent components on the fertilizing capacity of chicken semen stored at 5°C. *Poult. Sci.*, 57: 277-284.
5. Ngoula, F., T.T. Tebug, A. Kenfack, F.H. Defang, F. Tendonkeng and T.E. Pamo, 2012. Effects of buck age, storage duration, storage temperature and diluent on fresh west african dwarf buck semen. *J. Reprod. Infertil.*, 3: 58-66.
6. Habibullah, M., M.A. Hashem, M.S. Rana and M.H. Islam, 2015. Effect of artificial insemination on different production parameter in hubbard classic broiler parent stock. *J. Bangladesh Agric. Univ.*, 13: 71-77.
7. Brillard, J.P., 1993. Sperm storage and transport following natural mating and artificial insemination. *Poult. Sci.*, 72: 923-928.

8. Hudson, G.H., A.V. Omprakash and K. Premavalli, 2016. Effect of semen diluents and dilution rates on motility of guinea fowl spermatozoa under short-term storage. *Indian Vet. J.*, 93: 13-15.
9. Parker, H.M. and C.D. McDaniel, 2003. Semen dilution prior to analysis influences the ability of the sperm quality analyzer to predict fertility whether inseminating with a constant number of sperm or a constant volume of semen. *Poult. Sci.*, 82: 1808-1815.
10. Abouelezz, F.M.K., C. Castano, A. Toledano-Diaz, M.C. Estes, A. Lopez-Sebastian, J.L. Campo and J. Santiago-Moreno, 2015. Sperm-egg penetration assay assessment of the contraceptive effects of glycerol and egg yolk in rooster sperm diluents. *Theriogenology*, 83: 1541-1547.
11. Kuzlu, M. and A. Taskin, 2017. The effect of different extenders on the sperm motility and viability of frozen turkey semen. *Indian J. Anim. Res.*, 51: 235-241.
12. Rahman, M.S., M.R. Gofur, M.M. Rahman, F.Y. Bari and N.S. Juyena, 2018. Effect of skim milk and tris-citrate extenders to preserve the semen of indigenous ram of Bangladesh. *Asian J. Biol.*, 5: 1-11.
13. Al-Daraji, H.J., 2012. Effect of diluent supplementation with different levels of orange juice on semen quality during liquid storage of Roosters' semen. *Int. J. Vet. Sci.*, 1: 5-9.
14. El-Nasry, E., H.M. Khalil, M. Abaza and A. El-Saadany, 2004. Use of antioxidants in storing local cockerels semen. 1. Effects on semen quality and fertility. *Proceedings of the 22nd World's Poultry Congress*, June 8-13, 2004, Istanbul, Turkey, pp: 231-235.
15. Fakhriidin, M.B.M.R. and N.K. Flayyih, 2011. A new simple medium for *in vitro* sperm activation of asthenozoospermic patients using direct swim-up technique. *Kufa Med. J.*, 14: 67-75.
16. Evans, G. and W.M.C. Maxwell, 1987. *Salamon's Artificial Insemination of Sheep and Goats*. Butterworths Pvt. Ltd., Sydney, Australia, ISBN-13: 9780409491777, Pages 196.
17. Karabinus, D.S., D.P. Evenson and M.T. Kaproth, 1991. Effects of egg yolk-citrate and milk extenders on chromatin structure and viability of cryopreserved bull sperm. *J. Dairy Sci.*, 74: 3836-3848.
18. Blom, E. and N.O. Christensen, 1947. Studies on the pathological conditions in the testis, epididymis and accessory glands in the bull. *Skand. Vet. Tidsskr.*, 37: 1-49.
19. Chemineau, P., Y. Guerin, Y. Caginie, P. Arguer and J.C. Vallet, 1991. Training manual on artificial insemination in sheep and goats. *FAO Animal Production and Health Paper No. 83*, FAO, Rome, Italy.
20. Hancock, J.L., 1952. The morphology of bull spermatozoa. *J. Exp. Biol.*, 29: 445-453.
21. Swanson, E.W. and H.J. Bearden, 1951. An eosin-nigrosin stain for differentiating live and dead bovine spermatozoa. *J. Anim. Sci.*, 10: 981-987.
22. Sikka, S.C., 2004. Role of oxidative stress and antioxidants in andrology and assisted reproductive technology. *J. Androl.*, 25: 5-18.
23. Al-Taji, H.A., 1994. Sperm activation and intrauterine insemination: The effect of sperm concentration and culture media on sperm activation potential *in vitro*. M.Sc. Thesis, College of Medicine, University of Baghdad, Iraq.
24. Parks, J.E. and D.V. Lynch, 1992. Lipid composition and thermotropic phase behavior of boar, bull, stallion and rooster sperm membranes. *Cryobiology*, 29: 255-266.
25. Pillai, B.K., R. Jasuja, J.R. Simard and J.A. Hamilton, 2009. Fast diffusion of very long chain saturated fatty acids across a bilayer membrane and their rapid extraction by cyclodextrins: Implications for adrenoleukodystrophy. *J. Biol. Chem.*, 284: 33296-33304.
26. Brunaldi, K., N. Huang and J.A. Hamilton, 2010. Fatty acids are rapidly delivered to and extracted from membranes by methyl- β -cyclodextrin. *J. Lipid Res.*, 51: 120-131.
27. Bergeron, A., Y. Brindle, P. Blondin and P. Manjunath, 2007. Milk caseins decrease the binding of the major bovine seminal plasma proteins to sperm and prevent lipid loss from the sperm membrane during sperm storage. *Biol. Reprod.*, 77: 120-126.
28. Khaeim, H.M., 2013. Mass selection with an optical sorter for head scab resistance in soft red winter wheat. Master's Thesis, College of Agriculture, Food and Environment, University of Kentucky, Lexington, KY., USA.