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# **Research Article**

# Anticoccidial Effect of a *Negilla sativa* Seed-based Diet on *Eimeria tenella* Infection in Chickens

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# **Abstract**

**Background and Objective:** *Eimeria tenella* (*E. tenella*) infection is one of the most common and widely distributed parasitic diseases in chickens. Anticoccidial agents are used to treat the disease; however, due to the rise of drug-resistant strains and the ban of anticoccidials in many countries, alternative treatments are necessary. This study was conducted to analyze the effectiveness of *Nigella sativa* crushed seeds and ionophore salinomycin on *E. tenella* infection in broiler chickens. **Methodology:** One hundred one-day-old broiler chicks were divided into 4 equal groups. Group 1 (G1) was fed a basal anticoccidial-free diet containing 1% crushed *Nigella sativa* seeds and Group 2 (G2) was fed a basal diet containing 60 g t<sup>-1</sup> of salinomycin for 35 days. Groups 3 (G3) and Group 4 (G4) were fed the basal diet only. G1, G2 and G3 were challenged with 1×10<sup>4</sup> sporulated oocysts of *E. tenella* and G4 served as the unchallenged control group. **Results:** The results revealed significant increases in glutathione (GSH) concentration, glutathione peroxidase (GSH-Px), superoxide dismutase (SOD) and catalase (CAT) activities at days 28 and 35 in G1 compared to G3 group. Nevertheless, a significant decrease in plasma malondialdehyde (MDA) was found in G1, G2 and G4 following the *E. tenella* challenge. Furthermore, G1 had significantly minor intestinal lesions scores and lower oocyte shedding compared to G3. **Conclusion:** It is concluded that *Nigella sativa* can be used to prevent and monitor avian Coccidiosis.

Key words: Eimeria tenella, Nigella sativa, oxidative stress, broilers, anticoccidial

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

#### **INTRODUCTION**

Coccidiosis is one of the most economically important diseases affecting commercial poultry<sup>1</sup>. Coccidiosis is related to intracellular protozoan parasites of the genus Eimeria and causes disruption of the intestinal epithelium, leading to diminished feed efficiency and loss of body weight<sup>1-3</sup>. Currently, in intensively reared poultry settings, this infection is controlled by the use of anticoccidial medications and/or by vaccination<sup>4</sup>. However, due to the increase in medicationresistant strains of Eimeria and rising public distress about medication remain in chicken products, additional treatments are urgently required<sup>5,6</sup>. Several studies have investigated the use of various dietary supplements and probiotics to control Eimeria infections7-10. Nigella sativa and its active constituents have been found to exhibit antioxidant activity, as well as immunomodulatory, immunotherapeutic potentials, antitoxic, anti-histaminic, anti-inflammatory properties and anti-helmenthic effects<sup>11,12</sup>.

This study evaluated the effects of *Nigella sativa*-based diets as natural anticoccidials through their antioxidant behavior to control *E. tenella* infections in broiler chickens.

#### **MATERIALS AND METHODS**

After cleaning and disinfecting pens, 100 one-day-old broiler chicks (Ross 308) were randomly assigned to four equal groups of 25 chicks each. Group 1 (G1) was fed with a basal diet free of anticoccidial drug and supplemented with 1% whole crushed Nigella sativa seeds and Group 2 (G2) was fed with a basal diet containing salinomycin (Bio-Cox) at a rate of 60 g  $t^{-1}$  for 35 days. Group 3 (G3) and Group 4 (G4) were fed the basal diet only. G3 served as the challenged untreated control and G4 served as the untreated unchallenged control. G1, G2 and G3 groups were challenged with  $1 \times 10^4$  mature oocysts of E. tenella. Nigella sativa crushed seeds were mixed daily with the basal diet for the duration of the experiment. Feed and water were provided ad libitum and continuous 24 h of lighting was applied. The environmental temperature was decreased gradually from 33°C on day 1 to 24°C (±1) on day 21. Chickens were vaccinated against Newcastle disease, infectious bronchitis and infectious bursal disease. To determine gut lesion scores, 6 chickens were randomly chosen from each group for scoring at day 7 after the *Eimeria* challenge<sup>13</sup>.

*Eimeria* infection and assessment of fecal oocyst production: The oocysts of a field isolate of *E. tenella* were

obtained from a broiler farm that had chickens with bloody diarrhea, pathognomonic cecal lesions and a high mortality rate. Oocysts were collected from cecal contents and sporulated in a 2.5% potassium dichromate solution. Chickens in G1, G2 and G3 were orally challenged with  $1\times10^4$  sporulated oocysts of *E. tenella* at 28 days of age. Fecal materials were collected from the litter at 10 days post-infection and the numbers of oocysts  $g^{-1}$  were assessed using a hemocytometer counting chamber  $g^{-1}$ .

**Blood sampling:** Blood samples (2-3 mL) were taken from separate chickens (n = 6) at days 28 and 35 from the jugular vein and collected into tubes containing Ethylenediaminetetraacetic acid (EDTA) as an anticoagulant for the determination of glutathione (GSH) level, glutathione peroxidase (GSH-Px), superoxide dismutase (SOD) and catalase (CAT) activities in the hemolysate. Hemoglobin content (Hb) was performed as previously described by Jain<sup>15</sup>. Blood plasma was used for the estimation of malondialdehyde (MDA) as previously described by Ohkawa *et al.*<sup>16</sup>.

**Hemolysate preparation:** At 28 and 35 days of age, blood samples were collected in EDTA and centrifuged at 3000 rpm for 10 min; the buffy coat and plasma were separated from erythrocytes. Erythrocytes were washed three times with 0.9% normal saline and then 20% (v/v) hemolysate was prepared. The Beutler *et al.*<sup>17</sup> procedure was used to measure hemolysate activities levels of GSH. GSH-Px was measured according to the method of Paglia and Valentine<sup>18</sup>, SOD was measured according to the method of Marklund and Marklund<sup>19</sup> and CAT was measured according to the method of Aebi<sup>20</sup>.

**Statistical analysis:** The data were analyzed with SPSS 16.0 (SPSS Inc., Chicago, IL, USA) using one-way analysis of variance and least significance differences to determine the difference between groups at level of p<0.05.

#### **RESULTS**

Tables 1, 2, 3 and 4 show GSH levels, GSH-Px, SOD and CAT activities of the chickens at 28 and 35 days. From day 28, broilers from G1 group had the highest (p<0.05) GSH levels, GSH-Px, SOD and CAT activities. At day 35, G3 group had the lowest (p<0.05) GSH levels, as well as GSH-Px, SOD and CAT activities.

Significant levels of plasma MDA were apparent in G3 at day 35 only (Table 5). Nevertheless, plasma MDA levels in the

Table 1: Levels of glutathione (GSH) in erythrocyte hemolysate in the different groups of chickens at 28 and 35 days of age

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Parameter	Groups	28 days	35 days
GSH (nmol mL <sup>-1</sup> Hb)	G1	17.43±2.00a	13.61±1.04ª
	G2	13.33±0.98 <sup>b</sup>	10.86±0.92 <sup>b</sup>
	G3	12.90±1.30 <sup>b</sup>	8.23±1.15°
	G4	13.49±1.12 <sup>b</sup>	$12.94\pm0.80^{a}$

a,b,cp<0.05, SD: Standard deviation of means

Table 2: Levels of glutathione peroxidase (GSH-Px) activity in erythrocyte hemolysate in the different groups of chickens at 28 and 35 days of age

Parameter	Groups	28 days	35 days
GSH-Px (U g <sup>-1</sup> Hb)	G1	9.82±0.80 <sup>a</sup>	8.20±1.00°
	G2	6.64±1.47 <sup>b</sup>	5.57±1.28bc
	G3	7.78±1.11 <sup>b</sup>	4.50±1.56°
	G4	7.39±1.78 <sup>b</sup>	7.92±0.89ª

a,b,cp<0.05, SD: Standard deviation of means

Table 3: Levels of superoxide dismutase (SOD) activity in erythrocyte hemolysate in the different groups of chickens at 28 and 35 days of age

Parameter	Groups	28 days	35 days
SOD (U g <sup>-1</sup> Hb)	G1	49.29±2.24 <sup>a</sup>	43.37±5.30°
	G2	37.90±6.29b	36.29±4.39ab
	G3	39.13±4.30 <sup>b</sup>	30.55±6.00 <sup>b</sup>
	G4	40.28±3.75 <sup>b</sup>	41.30±3.28 <sup>a</sup>

a,bp<0.05, SD: Standard deviation of means

Table 4: Levels of catalase (CAT) activity in erythrocyte hemolysate in the different groups of chickens at 28 and 35 days of age

Parameter	Groups	28 days	35 days
CAT (U g <sup>-1</sup> Hb)	G1	60.25±4.64 <sup>a</sup>	56.38±6.35°
	G2	49.99±3.38b	45.31±5.93b
	G3	47.31±5.39 <sup>b</sup>	32.11±7.32°
	G4	50.00±4.25 <sup>b</sup>	53.29±4.44°

a,b,cp<0.05, SD: Standard deviation of means

Table 5: Levels of malondialdehyde (MDA) in blood plasma in the different groups of chickens at 28 and 35 days of age

Parameter	Groups	28 days	35 days
MDA (nmol mL <sup>-1</sup> plasma)	G1	3.851±0.282a	5.358±0.475b
	G2	$4.350\pm0.350^{a}$	5.981±0.501 <sup>b</sup>
	G3	$3.909 \pm 0.218^a$	$7.870 \pm 0.600^a$
	G4	$4.089 \pm 0.118^a$	4.126±0.217°

a,b,cp<0.05, SD: Standard deviation of means

G1 and G2 groups were comparable. No significant differences were observed between all experimental groups at day 28.

The result of oocyst output at 28 and 35 days of age (10 days post-challenge infection) are presented in Fig. 1. The G3 group had the highest mean numbers of oocysts, with approximately  $2.31\times10^6$  total oocysts shed. The G1 and G2 groups shed approximately  $0.41\times10^6$  and  $0.34\times10^6$  total oocysts, respectively.

The lesion scores of broilers during the experimental period (mean±SD) are shown in Fig. 2. Lesion scores were zero at 28 days for all the groups before the challenge. After the challenge, the G3 group had the highest mean lesion

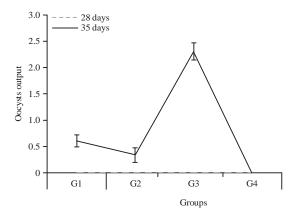


Fig. 1: Oocysts output ( $\times 10^6$ ) in the different groups of chickens at 28 and 35 days of age

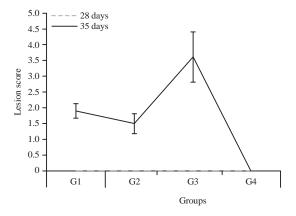


Fig. 2: Lesion scores in the different groups of chickens at 28 and 35 days of age

scores with an average of 3.6, while the mean lesions scores for the G1 and G2 groups were 1.8 and 1.6, respectively.

#### **DISCUSSION**

Reactive oxygen species (ROS) are molecules that are normally produced inside cells and tissues. The effects of ROS are usually mitigated by internal antioxidants and repair systems; however, the negative effects of ROS are not completely resolved and some of these effects accumulate over time to cause oxidative stress, a condition in which ROS production is greater than internal antioxidant systems<sup>21</sup>. Consequently, oxidative damage to different types of macromolecules can occur, which may be involved in the pathogenesis of many different diseases<sup>21,22</sup>.

Results from this study showed that supplementation with *Nigella sativa* seeds in the diet significantly increased levels of different antioxidants on day 28 (GSH, GSH-Px, SOD and CAT). The antioxidant properties of *Nigella sativa* seeds have

been reported in many other previous studies<sup>23</sup>. It is found that up-regulation of these antioxidants in G1 was paralleled by a reduction in oxidative damage (MDA). These results are consistent with previous studies in which the oxidative damage (MDA) was significantly mitigated by enzymatic and non-enzymatic antioxidants<sup>24</sup>.

It is also found that the markers of antioxidants measured on day 35 were significantly higher in G1 compared to the other infected groups (G2 and G3), which led to decreased levels of oxidative damage (MDA) in both groups (G1 and G2). G3 had the lowest levels of antioxidants and highest levels of oxidative damage. These results were directly associated with the significant changes in the degree of lesions and the oocyte outputs among groups. G3 had the highest levels of lesions and oocyte outputs, while G2 and G1 had lower levels of lesions and oocyte outputs. The findings here highlight the effectiveness of herbal supplementation and the role of antioxidant properties of *Nigella sativa* seeds in recovery from parasitic infection. These seeds may be used in the diet instead of commercial chemicals like salinomycin.

#### CONCLUSION

*Nigella sativa* has an antioxidant capacity that can boost defense mechanisms of chickens against infection. This study recommends that *Nigella sativa* can be used commercially in the diet as a protective agent.

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#### REFERENCES

- 1. McDonald, V. and M.W. Shirley, 2009. Past and future: Vaccination against *Eimeria*. Parasitology, 136: 1477-1489.
- 2. Morris, G.M. and RB Gasser, 2006. Biotechnological advances in the diagnosis of avian coccidiosis and the analysis of genetic variation in *Eimeria*. Biotechnol. Adv., 24: 590-603.
- 3. Dalloul, R.A. and H.S. Lillehoj, 2005. Recent advances in immunomodulation and vaccination strategies against coccidiosis. Avian Dis., 49: 1-8.
- 4. Quiroz-Castaneda, R.E. and E. Dantan-Gonzalez, 2015. Control of avian coccidiosis: Future and present natural alternatives. BioMed. Res. Int., Vol. 2015. 10.1155/2015/430610.

- 5. Williams, R.B., 2006. Tracing the emergence of drug-resistance in coccidia (*Eimeria* spp.) of commercial broiler flocks medicated with decoquinate for the first time in the United Kingdom. Vet. Parasitol., 135: 1-14.
- Bafundo, K.W., H.M. Cervantes and G.F. Mathis, 2008. Sensitivity of *Eimeria* field isolates in the united states: Responses of nicarbazin-containing anticoccidials. Poult. Sci., 87: 1760-1767.
- Jang, S.I., M.H. Jun, H.S. Lillehoj, R.A. Dalloul, I.K. Kong, S. Kim and W. Min, 2007. Anticoccidial effect of green tea-based diets against *Eimeria maxima*. Vet. Parasitol., 144: 172-175.
- 8. Molan, A.L., Z. Liu and S. De, 2009. Effect of pine bark (*Pinus radiata*) extracts on sporulation of coccidian oocysts. Folia Parasitol., 56: 1-5.
- 9. Nweze, N.E. and I.S. Obiwulu, 2009. Anticoccidial effects of *Ageratum conyzoides*. J. Ethnopharmacol., 122: 6-9.
- 10. Anosa, G.N. and O.J. Okoro, 2011. Anticoccidial activity of the methanolic extract of *Musa paradisiaca* root in chickens. Trop. Anim. Health Prod., 43: 245-248.
- 11. Leong, X.F., M.R. Mustafa and K. Jaarin, 2013. *Nigella sativa* and its protective role in oxidative stress and hypertension. Evidence-Based Complement. Altern. Med., Vol. 2013. 10.1155/2013/120732.
- 12. Salem, M.L., 2005. Immunomodulatory and therapeutic properties of the *Nigella sativa* L. seed. Int. Immunopharmacol., 5: 1749-1770.
- 13. Johnson, J. and W.M. Reid, 1970. Anticoccidial drugs: Lesion scoring techniques in battery and floor-pen experiments with chickens. Exp. Parasitol., 28: 30-36.
- Hodgson, J.N., 1970. Coccidiosis: Oocyst counting technique for coccidiostat evaluation. Exp. Parasitol., 28: 99-102.
- 15. Jain, N.C., 1993. Essentials of Veterinary Hematology. Lea and Febiger, Philadelphia, PA., USA., ISBN-13: 9780812114379, Pages: 417.
- 16. Ohkawa, H., N. Ohishi and K. Yagi, 1979. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal. Biochem., 95: 351-358.
- 17. Beutler, E., O. Duron and B.M. Kelly, 1963. Improved method for the determination of blood glutathione. J. Lab. Clin. Med., 61: 882-888.
- 18. Paglia, D.E. and W.N. Valentine, 1967. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. J. Lab. Clin. Med., 70: 158-169.
- 19. Marklund, S. and G. Marklund, 1974. Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. Eur. J. Biochem., 47: 469-474.

- 20. Aebi, H., 1984. Catalase *in vitro*. Methods Enzymol., 105: 121-126.
- 21. Halliwell, B. and J.M.C. Gutteridge, 2007. Free Radicals in Biology and Medicine. 4th Edn., Clarendon Press, Oxford, UK., ISBN-13: 9780198568698, Pages: 888.
- 22. Pamplona, R. and D. Costantini, 2011. Molecular and structural antioxidant defenses against oxidative stress in animals. Am. J. Physiol. Regul. Integr. Comp. Physiol., 301: R843-R863.
- 23. Sogut, B., I. Celik and Y. Tuluce, 2008. The effects of diet supplemented with the black cumin (*Nigella sativa* L.) upon immune potential and antioxidant marker enzymes and lipid peroxidation in broiler chicks. J. Anim. Vet. Adv., 7: 1196-1199.
- 24. Abd Elgawad, H., G. Zinta, M.M. Hegab, R. Pandey, H. Asard and W. Abuelsoud, 2016. High salinity induces different oxidative stress and antioxidant responses in maize seedlings organs. Front. Plant Sci., Vol. 7. 10.3389/fpls.2016.00276.