ISSN 1682-8356 ansinet.org/ijps



# POULTRY SCIENCE

ANSImet

308 Lasani Town, Sargodha Road, Faisalabad - Pakistan Mob: +92 300 3008585, Fax: +92 41 8815544 E-mail: editorijps@gmail.com

# Plasma Selenium Concentration and Glutathione Peroxidase Activity in Red Blood Cells of Laying Hens Fed Sodium Selenite or Zinc-L-Selenomethionine

A. Chantiratikul<sup>1\*</sup>, W. Aengwanich<sup>1</sup>, O. Chinrasri<sup>1</sup> and P. Chantiratikul<sup>2</sup>
<sup>1</sup>Faculty of Veterinary Medicine and Animal Science, Mahasarakham University,
Muang, Maha Sarakham 44000, Thailand
<sup>2</sup>Department of Chemistry, Faculty of Science, Mahasarakham University,
Kantarawichai, Maha Sarakham 44150, Thailand

Abstract: The objective of this study was to determine the effect of sodium selenite and zinc-L-selenomethionine on plasma selenium (Se) concentration and glutathione peroxidase (GSH-Px) activity in the red blood cells (RBC) of laying hens. Two hundred twenty four CP Browns aged 71 weeks were divided according to a 2x3 factorial in completely randomized design. One more group without additional Se supplementation was used as negative control. Each treatment consisted of four replicates and each replicate contained eight hens. The dietary treatments were T1: basal diet, T2, T3 and T4: basal diets added 0.3, 1.0 and 3.0 mg Se from sodium selenite/kg, respectively, T5, T6 and T7: basal diets added 0.3, 1.0 and 3.0 mg Se from zinc-L-selenomethionine/kg, respectively. The findings revealed that the plasma Se concentration of hens received supplemented zinc-L-selenomethionine diets was higher (p<0.05) than that of hens received supplemented sodium selenite diets. Plasma Se concentration statistically increased (p<0.01) with increasing Se levels. The Se sources did not dramatically alter GSH-Px activity in RBC. However, GSH-Px activity significantly increased (p<0.01) with increasing dietary selenium levels. The results in this experiment indicate that zinc-L-selenomethionine increases higher plasma Se concentration than sodium selenite, however GSH-Px activity in RBC of laying hens is not affected by Se sources.

**Key words:** Sodium selenite, zinc-L-selenomethionine, plasma selenium concentration, glutathione peroxidase activity, laying hens.

## Introduction

Selenium is an essential nutrient for poultry. N.R.C. (1994) recommended 0.05 mg Se/kg in the diet of laving hens for the maintenance and egg production. Moreover, Se allowance higher than 0.1 mg/kg can improve immunity (Song et al., 2006). Selenium is an important component of the antioxidant defense mechanism and functions by controlling the body's glutathione pool and its major Se-containing antioxidant enzyme, GSH-Px (Mahmoud and Edens, 2003). The main physiological function of GSH-Px is to maintain low levels of H2O2 and others hydroperoxides in the cell to prevent tissues from peroxidation damages (Kim and Mahan, 2003). Recent studies found that activity of blood GSH-Px in chicks fed organic Se in the form of Se-enriched yeast and sodium selenite was not different (Kuricova et al., 2003; Payne and Southern, 2005). On the other hand, Mahmoud and Edens (2003) reported that Se-enriched yeast had elevated higher GSH-Px activity in blood of chickens in a thermoneutral environment and after heat distress than inorganic Se. Nevertheless, GSH-Px activity increased significantly (P<0.05) with increasing supplementation in the diet of broilers (Omaye and Tappel, 1974; Kuricova et al., 2003; Yoon et al., 2007) and laying hens (Payne and Southern, 2005). Furthermore, plasma Se concentration of chickens fed

Se-enriched yeast increased significantly (P<0.05) when compared that of chickens fed inorganic Se (Kuricova et al., 2003; Payne and Southern, 2005). The above results indicated that Se-enriched yeast was more beneficial than inorganic Se. However, there is lack information of the use of zinc-L-selenomethionine in laying hens. Zinc-L-selenomethionine is designed to be highly soluble and increase bioavailability of selenium (Ward, 2003). The previous studies showed that zinc-Lselenomethionine higher improved Se status in horses (Richardson et al., 2006) and increased muscle (George et al., 2004) and plasma Se concentrations in broilers (Spears et al., 2003) than sodium selenite. Thus, the aim of this study was to determine the effect of zinc-Lselenomethioine and sodium selenite on plasma Se concentration and GSH-Px activity in RBC of laying hens.

# **Materials and Methods**

Two hundred and twenty four CP Brown laying hens, 71 weeks old, were housed in evaporative cooling system housing. Internal temperature was set at 24°C. Lights were on continuously. The hens were randomly divided into 7 groups. Each group consisted of 4 replicates with 8 hens per replicate. The basal diet (Table 1) was formulated to meet or exceed nutrient requirement according to N.R.C. (1994) and without Se

Table 1: Feed ingredient and chemical composition of basal

ulet.	
Ingredients	%
Corn	59.00
Rice bran	4.25
Soybean meal (44%CP)	16.00
Fish meal	6.36
Soybean oil	2.78
Dicalcium phosphate	1.65
Oyster shell meal	8.44
DL-Methionine	0.15
Salt	1.12
Vitamin-mineral premix <sup>2</sup>	0.25
Analyzed chemical composition (%DM)	
Dry matter	91.37
Crude protein	15.86
Ether extract	3.52
Crude fiber	2.87
Ash	12.85
ME³, kcal/kg	2,950.12

<sup>1</sup>Sodium selenite and zinc-L-selenomethionine were mixed in corn and added to the diet to achieve the treatment levels.

 $^2$ Vitamin-mineral premix provide (per kg diet): 10,000 IU vitamin A, 2,000 IU vitamin D<sub>3</sub>, 11mg vitamin E, 1.5 mg vitamin K ,  $_3$ 1.5 mg thiamin, 4 mg riboflavin, 10 mg pantothenic acid, 0.4, folic acid, 4 mg pyridoxine, 22 mg niacin, 0.4 mg colabamin, 0.1 mg biotin, 60 mg Fe, 70 mg, Mn, 50 mg Zn, 8 mg Cu, 0.5 mg Co, 0.7 mg I.  $^3$ Calculated value.

supplementation. The 0.3, 1.0, and 3.0 mg Se/kg from sodium selenite or zinc-L-selenomethionine (Availa®Se, Zinpro Corporation) were supplemented to the basal diet. Total Se concentration of zinc-L-selenomethionine was 1000 mg Se/kg. The hens received the basal diet or Se supplemented diets and water ad libitum throughout the 6 weeks experimental period. The experimental diets were randomly collected at the end of each week and determined for chemical composition (AOAC, 1999) and Se content. The same 2 hens in each replicate of the treatments were bled at the beginning of the experiment and the end of each tested week. Blood samples were collected via the main wing vein and placed into 10-ml tubes containing EDTA. After collection, the blood samples were centrifuged at 3,000 x g for 10 min. The plasma was harvested and stored at -20°C prior to determination of Se concentration. The dietary and plasma samples were digested with nitric acid until the solution was cleared. Selenium was analyzed by inductively coupled plasma mass spectrometer (ICP-MS model Elan-e, Perkin-Elmer SCIEX, USA) following the procedure of Sieniawska et al. (1999). The activity of GSH-Px in RBC was investigated on weeks 0, 1, 3 and 5. After separating plasma, RBC were washed three times with 0.9% saline solution. The distilled water was added to RBC to make a ratio of 4:1 and frozen for 24h to hemolyze the RBC (Patricia et al., 1982). The activity of GSH-Px in RBC was determined using the proposed procedure of Koller et al. (1984).

**Statistical analysis:** The data of plasma Se concentration and activity of GSH-Px in RBC were analyzed using GLM procedure appropriate for Factorial in Completely Randomized Design (SAS, 1996). Treatment differences were determined by orthogonal contrasts

- (1) basal diet vs. Se supplemented diets,
- (2) sodium selenite vs. zinc-L-selenomethionine,
- (3) levels of Se supplementation. Values of P<0.05 were considered significant.

#### **Results and Discussion**

The Se determination in experimental diets indicated that the basal diet provided 0.30 mg Se/kg. Consequently, the basal diets added 0.3, 1.0 and 3.0 mg Se/kg from sodium selenite or zinc-L-selenomethionine contained 0.68, 1.14 and 3.37, or 0.77, 1.43 and 3.47 mgSe/kg, respectively. Plasma Se concentration of laying hens fed control diet was lower (P<0.05) than that of hens fed Se supplemented diets (Table 2). Zinc-Lselenomethionine increased higher (P<0.05) plasma Se concentration than sodium selenite since the first week of the trial. The results in this experiment are in agreement with the previous reports (Kuricova et al., 2003; Spears et al., 2003; Payne and Southern, 2005; Yoon et al., 2007). Those reports found blood Se concentration of birds receiving organic Se in the forms of zinc-L-selenomethionine or Se-enriched yeast was significantly higher (P<0.05) than that of birds kept on basic diet or diet supplemented with sodium selenite. Normally, organic Se in the form of selenomethionine increased blood Se levels to a much greater extent than did selenite (Thomson, 1998) or selenate (Beilstein and Whanger, 1986). On the other hand, Pan et al. (2007) observed whole-blood Se concentrations of hens fed sodium selenite were higher (P<0.05) than those of hens fed Se-enriched yeast. That study explained that organic Se is mainly deposited in egg and body tissue, whereas inorganic Se remains in the blood. Plasma Se concentration increased significantly (P<0.05) with the increment of dietary Se, regardless of Se source (Table Similarly, Jiakui and Xiaolong (2004) and Zuberbuehler et al. (2006) observed plasma Se of laying hens increased dramatically (P<0.05) with levels of Se supplementation. The significant differences between the effects of inorganic and organic selenium on plasma Se showed since the first week of the present experiment. However, Kuricova et al. (2003) found those effects after six weeks. This discrepancy in results may be due to the differences of Se content in the basal diet and concentration of Se used. The results demonstrated that GSH-Px activity in RBC of laying hens fed control diet was lower (P<0.05) than that of hens fed Se supplemented diets. Sources of Se did not markedly alter (P>0.05) GSH-Px activity in RBC of laying hens (Table 3). The outcome was in agreement with the results of Omaye and Tappel (1974), Cantor et al. (1975), Kuricova et al. (2003); Payne and Southern

Table 2: Plasma selenium concentrations (µg/ml) of laying hens fed sodium selenite or zinc-L-selenomethionine

Experi-	Basal Diet	Sodium selenite (mg/kg)			Zinc-L-s	P-value <sup>1</sup>						
mental week												
		0.3	1.0	3.0	0.3	1.0	3.0	SEM	В	S	L	SxL
0	0.13	0.14	0.13	0.14	0.13	0.13	0.14	0.00	NS	NS	NS	NS
1	0.10	0.14	0.18	0.25	0.15	0.23	0.41	0.02	*	*	*	*
2	0.10	0.16	0.19	0.25	0.21	0.27	0.53	0.02	*	*	*	*
3	0.10	0.17	0.18	0.31	0.20	0.35	0.67	0.03	*	*	*	*
4	0.11	0.16	0.18	0.23	0.18	0.28	0.52	0.02	*	*	*	*
5	0.14	0.19	0.22	0.31	0.22	0.34	0.71	0.03	*	*	*	*
6	0.12	0.12	0.23	0.32	0.17	0.27	0.48	0.02	*	*	*	*
Average	0.11	0.15	0.18	0.26	0.18	0.27	0.49	0.00	*	*	*	*

<sup>&</sup>lt;sup>1</sup>B = basal diet ∨s. others, S = sodium selenite ∨s. zinc-L-selenomethionine, L = levels of Se supplementation, SxL = Se sources x levels. \* = significantly difference at P<0.05. NS = not significantly difference at P<0.05.

Table 3: Glutathione peroxidase activity in red blood cell (mUnit/Hb) of laying hens fed sodium selenite or zinc-L-selenomethionine

Experi-		Sodium selenite (mg/kg)			Zinc-L-selenomethionine (mg/kg)				P-value <sup>1</sup>			
mental	Basal											
week	diet	0.3	1.0	3.0	0.3	1.0	3.0	SEM	В	S	L	SxL
0	2.68	5.76	3.33	6.99	4.13	4.04	4.46	0.39	NS	NS	NS	NS
1	4.76	8.75	14.70	15.49	11.60	14.03	16.05	0.94	*	NS	*	*
3	5.10	9.11	10.60	11.97	7.90	10.74	13.24	0.79	*	NS	*	NS
5	3.27	7.58	10.27	11.90	4.04	11.51	12.13	0.72	*	NS	*	NS
A∨erage	4.00	7.78	9.65	11.58	8.03	10.02	11.52	0.46	*	NS	*	NS

<sup>&</sup>lt;sup>1</sup>B = basal diet ∨s. others, S = sodium selenite ∨s. zinc-L-selenomethionine, L = levels of Se supplementation, SxL = Se sources x levels. \* = significantly difference at P<0.05. NS = not significantly difference at P>0.05.

(2005) and Yoon et al. (2007). The current results reflected that bioavailability of zinc-L-selenomethionine and sodium selenite with respect to GSH-Px activity in RBC was similar. This might be explained by their metabolic route. Selenomethionine is converted into selenocysteine that can be degraded further in liver to serine and selenide. Sodium selenite is converted initially to selenoglutathione trisulfide and then degraded in liver to form selenide. The selenide is finally used for selenoprotein synthesis, such as GSH-Px (Schrauzer, 2000). The activity of GSH-Px was normally dependent on the amount of Se supplied to the diet (Omaye and Tappel, 1974; Kuricova et al., 2003). Subsequently, the present result found that GSH-Px activity in RBC increased (P<0.05) accordingly with levels of Se supplementation (Table 3). From the obtained results, it could be concluded that zinc-L-selenomethionine (P<0.05) increased significantly plasma concentration. However, zinc-L-selenomethionine and sodium selenite were equally effective in stimulating GSH-Px activity in RBC of laying hens.

### Acknowledgments

This work was supported by a grant from Masasarakham University. Zinc-L-selenomethionine (Availa®Se) was provided by Zinpro Animal Nutrition (Thailand), Inc. The authors wish to thank Mr. Witpol Thosaikham, Miss. Galyanee Kunchree and Mr. Jakrit Charoensin for data collection and laboratory assistance.

#### References

AOAC, 1999. Official Methods of Analysis. 16<sup>th</sup> ed. Association of Official Analysis Chemists, Washington, D.C.

Beilstein, M.A. and P.D. Whanger, 1986. Deposition of dietary organic and inorganic selenium in rat erythrocyte proteins. J. Nutr., 166: 1701-1710.

Cantor, A.H., M.L. Scott and T. Noguchi, 1975. Biological availability of Selenium in feedstuffs and selenium compounds for prevention of exudative diathesis in chicks. J. Nutr., 105: 96-105.

George, B., S. Davis and T.L. Ward, 2004. A novel organic selenium source (zinc-L-selenomethionine, Availa® Se) for broilers. J. Anim. Sci., 82 (Suppl. 1): 319.

Jiakui, L. and W. Xiaolong, 2004. Effect of dietary organic versus inorganic selenium in laying hens on the productivity, selenium distribution in egg and selenium content in blood, liver and kidney. J. Trace Element Med. Biol., 18: 65-68.

Kim, Y.Y. and D.C. Mahan, 2003. Biological aspects of selenium in farm animals. Asian-Aust. J. Anim. Sci., 16: 435-444.

Koller, L.D., P.J. South., J.H. Exon., G.A. Whitbeck and J. Maas, 1984. Comparison of selenium levels and glutathione peroxidase activity in bovine whole blood. Can. J. Comp. Med., 48: 431-433.

Kuricova, S., K. Boldizarova., L. Gresakova., M. Levkut and L. Leng, 2003. Chicken selenium status when fed a diet supplemention with Se-Yeast, Actd. Vet., 72: 339-346.

- Mahmoud, K. and F.W. Edens, 2003. Influence of selenium sources on age-related and mild heat stress-related changes of blood and liver glutathione redox cycle in broiler chickens (*Gallus domesticus*). Comp. Biochem. Physiol. Part B., 136: 921-934.
- National Research Council, 1994. Nutrition Requirement of Poultry, 9<sup>th</sup> ed. National Academy Press, Washington, D.C. U.S.A.
- Omaye, S.T. and A.L. Tappel, 1974. Effect of dietary selenium on glutathione peroxidase in the chick. J. Nutr., 104: 747-753.
- Pan, C., K. Huang, Y. Zhao, S. Qin. F. Chen and Q. Hu, 2007. Effect of selenium source and level in hen's diet on tissue selenium deposition and egg selenium concentration. J. Agric. Food Chem., 55: 1027-1032.
- Partricia, A.P., A. Munyani and J. Beachum, 1982. Determination of selenium concentration and glutathione peroxidase activity in plasma and erythrocytes. Clin. Chem., 28: 311-316.
- Payne, R.L. and L.L. Southern, 2005. Comparison of inorganic and organic selenium sources for broilers. Poult. Sci., 84: 898-902.
- Richardson, S.M., P.D. Siciliano, T.E. Engle, C.K. Larson and T.L. Ward, 2006. Effect of selenium supplementation and source on the selenium status of horses. J. Anim. Sci., 84: 1742-1748.
- SAS, 1996. SAS/STAT® User's Guide (Release 6.03 ed.). SAS Inst. Inc. Cary, N.C.

- Schrauzer, G.N., 2000. Selenomethionine: a review of its nutritional significance, metabolism and toxicity. J. Nutr., 130: 1653-1656.
- Sieniawska, C.E., R. Mensikov and H.T. Delves, 1999. Determination of total selenium in serum, whole blood and erythrocytes by ICP-MS. J. Anal. At. Spectrom., 14: 109-112.
- Song, Z., Y. Guo and J. Yuan, 2006. Effects of dietary iodine and selenium on the activities of blood lymphocytes in laying hens. Asian-Aust. J. Anim. Sci., 19: 713-719.
- Spears, J.W., J. Grimes, K. Lloyd and T.L. Ward, 2003. Efficacy of a novel organic selenium compound (zinc-l-selenomethionine, Availa Se) in broiler chicks. Page: 197-198 in Proceeding of the 1<sup>st</sup> Latin Am. Congress of Ani. Nutr., Cancun, Maxico.
- Thomson, C.D., 1998. Selenium speciation in human body fluids. Analyst., 123: 827-831.
- Ward, T., 2003. Looking beyond selenium yeast. Feed Tech., 7: 20-22.
- Yoon, I., T.M. Werner and J.M. Butler, 2007. Effect of source and concentration of selenium on growth performance and selenium retention in broiler chickens. Poult. Sci., 86: 727-730.
- Zuberbuehler, C.A., R.E. Messikommer., M.M. Arnold., R.S. Forrer and C. Wenk, 2006. Effect of selenium depletion and selenium repletion by choice feeding on selenium status of young and old laying hens. Physiol. Behav., 87: 430-440.